Extraction and biological evaluation of Mycobacterium kansasii extracellular vesicles as a vaccine candidate against mycobacterial pulmonary infections

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ABSTRACT

Introduction: Extracellular vesicles (EVs) are bacterial products with diverse biological roles. Like many microorganisms, Mycobacterium kansasii as a nontuberculous mycobacteria (NTM), can naturally release EVs. The aim of the present study was the extraction and biological evaluation of M. kansasii as a vaccine candidate against mycobacterial pulmonary infections. Methods: After bacterial culture of the standard species of M. kansasii, the EVs extraction was done by density gradient ultracentrifugation method and biological evaluations of EVs were performed by SDS-PAGE and electron microscopy. Endotoxin safety of the EVs was evaluated by LAL test. Results: SDS-PAGE result showed more than 5 prominent protein bands (60-180 kDa). The intactness of the vesicles was verified by electron microscopy through which the spherical configuration of EVs with a diameter of 200-300 nm could be observed. The amount of lipopolysaccharide (LPS) contamination existing in EVs was in the specified application range of biological products. Conclusion: EVs were prepared with acceptable quality composition with intact conformational structure throughout the extraction procedure. The extracted EVs had the initial requirements as an immunogenic molecule, such as safety, stability, inexpensiveness and antigens possession which based on the similarities between M. kansasii and M. tuberculosis, make them a suitable candidate for future prophylactics, therapeutic, detection and adjuvants studies against mycobacterial pulmonary infections.

KEYWORDS: Extracellular vesicles (EVs), Mycobacterium kansasii, adjuvant.

INTRODUCTION

Previous studies have shown that bacteria release extracellular vesicles (EVs) naturally, also known as, outer membrane vesicles and membrane vesicles, under a variety of growth situations [1]. EVs are accounted for distinctive features such as, interaction of the microorganism with the environment, as well as the microbe’s physiology and pathogenesis, enzymes and toxins delivery, communication signals, presentation of recognized antigens to innate and adaptive immune system, defense against other microorganisms, resistance to host immune system, nutrition and survival, transfer of certain genes and biofilm formation [2-4]. EVs diverse biological roles, probably make them one of the bacterial evolution factors. Furthermore, EVs can be considered as a non-classical secretory system [5]. Considering the EVs capability to concentrate toxins and immune-modulatory molecules, the production of EVs is related to bacterial virulence. In this manner, the production of mycobacterial vesicles as a delivery mechanism of immunologically active molecules can lead to mycobacterial virulence which can be exploited to diagnose the pathogenic mycobacteria and the development of vaccines against them [6]. Like many other microorganisms, Mycobacterium (M.) kansasii naturally releases EVs in a variety of growth media. M. kansasii is the second most common cause of nontuberculous mycobacteria (NTM) infections [7, 8]. Although more than 150 different species of NTM have so far been identified, M. avium complex (MAC), M kansasii and M. abscessus are the causative agents of most pulmonary infections [9]. Buher and Pollak identified M. kansasii for the first time in 1953 and named it “yellow bacillus”. M. kansasii antigenic and clinical characteristics make it the most similar Mycobacterium to M. tuberculosis [10]. Due to the prevalence of NTM infections, studies to develop awareness about these microorganisms are important in order to prevent the disease from spreading among the high-risk individuals such as HIV patients. Mycobacterial extracts have been widely used in vaccines and adjuvants, such as Freund’s complete adjuvant. Mycobacterial cell wall contains a variety of antigens and the immune stimulating biomolecules, such as peptidoglycan, arabinogalactan, mycolic acids, phosphatidylinositol mannosides, tiocerol, lipomannan, and lipoarabinomann which can activate the dendritic cells. All
these compounds are effective molecular factors in the process of tuberculosis in mice and can induce immune responses against the infections. EVs derived from non-pathogenic mycobacteria such as M. bovis and M. smegmatis show high level of consistency with M. tuberculosis antigens. These remarkable property makes the EVs a suitable candidate for the next generation of vaccines or adjuvants [11]. Proteomic analyses of M. tuberculosis EVs have revealed approximately 50 proteins related to the virulence of the bacterium. These observations show that EVs can be used as an alternative to the active immunological M. tuberculosis virulence factors and could be used as a biomarker. Furthermore, it could be potentially used as a vaccine with no need to adjuvants [1]. The aim of the present study was the extraction and biological evaluation of M. kansasii EVs as a vaccine candidate.

MATERIALS and METHODS

Bacteria strain and culture condition

Standard strain of M. kansasii (CRBIP 7.42) was obtained from the standard bacterial collection of Mycobacteriology and Pulmonary Research Department, Pasteur Institute of Iran. The bacterial culture was initially grown in Lownenstein–Jensen medium (Pasteur Institute of Iran, Tehran, Iran) in order to activate the standard strain. After early preparation, the bacteria adjusted to 1 McFarland turbidity were cultured in 150 ml of Middlebrook 7H9 broth medium (Sigma, USA) at 37± 1°C for 4 weeks. The biomass was then harvested and the wet weight was evaluated.

EVs extraction

The EVs extraction and purification were performed by density gradient ultra-centrifugation method (Claassen’s method), using 3 extraction solutions. Firstly, 0.1 M Tris-HCl (Sigma, USA), pH 8.6± 0.1 and 10 mM EDTA (Sigma, UK) solution, secondly, 0.1 M Tris-HCl, pH 8.6± 0.1 and 10 mM EDTA and 100% (w/v) deoxycholate (Merck, Darmstadt, Germany) solution and thirdly, 0.1 M Tris-HCl, pH 8.6± 0.1 and 10 mM EDTA and 0.5% w/v deoxycholate solution [12, 13].

After deactivation of the bacterium for 2 h at 80°C, the cells were harvested by centrifugation at 1000 x g twice for 30 min. Then the supernatant was suspended in normal saline for 30 min and centrifuged at 1000 x g for 30 min. The bacterial biomass was then harvested in the first solution at 7.5 times of the wet weight for 30 min and then it was severely re-suspended by adding 1/20th volume of the second solution for 10 min. Finally, the suspension was centrifuged at 16000 x g at 4°C for 90 min. The obtained supernatant containing the EVs was concentrated by ultracentrifugation at 60000 x g at 4°C for 120 min and at 48000 x g at 4°C for 180 min. The EVs pellet was re-suspended in the third solution and was centrifuged at 60000 x g at 4°C for 120 min. The final pellet of EVs was re-suspended in 3% sucrose solution.

Physiochemical analysis

Protein assay

After extraction of EVs from 1.52 g harvested bacterial biomass, the total protein output was determined by Nano-drop technique to be 1.4 mg/ml. This result was also confirmed by Bradford protein assay.

SDS-PAGE

According to the SDS-PAGE result, we had more than 5 prominent protein bands with Mw between 60-180 kDa (Fig. 1).

Electron microscopy

The intactness of the vesicles was evaluated by SEM, and the EVs configuration revealed spherical bodies with a diameter of 180-4160 nm (Fig. 2).

LAL assay

The amount of lipopolysaccharide (LPS) contamination existing in EVs was under 300 IU, which was within the specified application range of the biological products.

RESULTS

Physiochemical analysis

Protein assay

After extraction of EVs from 1.52 g harvested bacterial biomass, the total protein output was determined by Nano-drop technique to be 1.4 mg/ml. This result was also confirmed by Bradford protein assay.

SDS-PAGE

The protein ingredients of EVs was analyzed by SDS-PAGE in 12% gel (using a Mini-PROTEAN® Tetra Cell - Bio-Rad) and the gel was stained with Coomassie Brilliant Blue (CBB).[13]

Electron microscopy

The intactness of the vesicles was checked by Scanning Electron Microscopy (SEM). For this purpose, field emission scanning electron microscope was used (FE-SEM; HITACHI S-4160 model with 5 nm image resolution, 20-30000 x magnification and maximum accelerating voltage of 30 kV). First, a small amount of suspension sample was coated with gold and then was observed with the electron microscope.

The Limulus Amebocyte Lysate (LAL) assay

Endotoxin safety was evaluated by LAL test (Pierce® LAL Chromogenic Endotoxin Quantitation Kit, Thermo Fisher Scientific Co. USA; 88282), according to the manufacturer’s instructions [13].
with its biological, antigenic and clinical characteristics can be considered as a vaccine candidate against mycobacterial pulmonary infections.

Moreover, EVs are recently considered as new generation of adjuvants. Salmani and colleagues have pointed to the necessity of developing more efficient vaccines with less side effects with new and improved adjuvant and have proposed the use of EVs as an adjuvant [16]. Mycobacterial extractions including Freund's complete adjuvant have been widely used in producing vaccines. Upon further verifications and experiments, biological compounds like M. kansasii EVs, can also be suggested as a new adjuvant which may replace chemical compounds such as alum in future vaccine designs.

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CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

REFERENCES